ORIGINAL PAPER

# Effect of Operating Parameters on Oil and Phenolic Extraction Using Supercritical CO<sub>2</sub>

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Received: 13 August 2009/Revised: 14 April 2010/Accepted: 21 April 2010/Published online: 16 May 2010 © AOCS 2010

Abstract Supercritical fluid extraction (SFE) with carbon dioxide was used to extract oil from canola press cake. Different operating conditions, e.g. pressure, temperature, and co-solvent % were investigated to optimize extraction parameters to yield canola meal with <4% oil. The residual oil content in the extracted canola meal reduced to 2.1-2.9% in our experimental trials. Residues of the optimum conditions based on oil yield were compared for the total phenolic content and the main phenolic compounds. Sinapine (the choline ester of sinapic acid) was the major phenolic constituent in both the SFE and n-hexane extracted canola meals and press cakes. n-Hexane extracted residues showed the retention of the highest sinapic acid, sinapine, sinapoyl glucose and total phenolic contents (mg/g) while the SF-extracted residues showed the lowest values for these compounds.

**Keywords** Supercritical fluid extraction · Canola press cake · Extraction parameters · Oil yield · *n*-Hexane extraction · Phenolic content

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#### Introduction

Canola, developed from traditional rapeseed by Canadian plant breeders during the 1970s, is the largest oilseed crop in Canada [1]. The production of canola oil involves two main processes: mechanical pressing and extraction, and further processing to remove impurities. Canola press cake, the by-product of canola oil processing, is used as a high-protein feed ingredient in the poultry, swine, cattle and fish industries [2, 3]. Due to its high oil content, the press cake is further extracted using solvent extraction (typically *n*-hexane) to retrieve much of the residual oil.

In the cold pressing process, oil is primarily removed through mechanical pressing at a relatively low temperature (typically near 60 °C). The cold pressing process results in only partial recovery of oil from the seed which usually yields canola meal of about 15-25% in oil content; therefore, pressing of seeds is often followed by n-hexane extraction at higher temperatures to achieve an approximately 96% recovery efficiency [4]. However, extraction requires large amounts of solvent and is very timeconsuming [5]. Additional processing (desolventisation/ toasting stage) is needed with an operating temperature of 100–110 °C to evaporate the solvent (*n*-hexane) from the meal [6]. Some compounds in the oil may undergo thermal degradation under solvent extraction conditions at high temperatures [7]. There is an increasing demand for new extraction techniques, which have shorter extraction times, no chemical residues, and a safer operation environment, to replace *n*-hexane extraction.

Supercritical fluid extraction has received increasing attention over the past two decades, including its use as a viable alternative to solvent extraction from oilseeds [8-12]. Supercritical fluid extraction utilizes the

combination of temperature and pressure to alter certain fluids to become excellent solvents for particular solutes. A supercritical state for a solvent is achieved when the temperature and pressure of the solvent is raised above its critical points. A supercritical fluid has the characteristics of both gases and liquids.

Supercritical  $CO_2$  extraction of canola seeds and meals has been reported [11, 13–18]. Several researchers studied the supercritical extraction of compounds, such as tocopherols, from rapeseed and canola seed and meals [14, 15]. Other studies focused on the effect of supercritical extraction on extracted oil properties, such as, fatty acid composition, oxidative stability [11, 16–18].

Some recent studies involving supercritical CO<sub>2</sub> extraction of lipids focused on extracting oil from dried distillers grains [5], and grape seeds [19]. While there has been a considerable amount of investigation on supercritical extraction of oil from canola seed and meals, literature is lacking on optimizing extraction conditions, such as, temperature and pressure, to achieve maximum oil extraction from the substrate. The first objective of this work was to investigate the extraction of oil from canola press cake using supercritical CO<sub>2</sub> extraction under different operating conditions, to optimize extraction parameters to yield canola meal with <4% residual oil. The yield of supercritical carbon dioxide extraction was compared with a traditional solvent extraction method (n-hexane extraction). The second objective was to examine the retention of phenolic components in the residues under the optimum SFE conditions.

### **Experimental Procedures**

Canola Press Cake Sample Preparation

Samples of canola (Nexera<sup>TM</sup>) press cake were obtained from Dow AgroSciences (Indianapolis, IN). Press cake was ground using a Retsch PM100 Planetary Ball Mill running at 450 RPM for 5 min to reduce and homogenize the particle size to an average of 473.9  $\mu$ m. The ground sample was then dried in a freeze dryer (FreeZone Plus 6 Liter Cascade Console Freeze Dry System, Lanconco) at -54 °C and 0.030 mBar for 48 h to bring down the moisture content from 6.29 to 1.64%. The dried sample was then used as the starting material for the supercritical fluid extraction (SFE) and *n*-hexane extraction. The sample was kept at -20 °C in the dark before use for extraction and analysis.

Supercritical Fluid Extraction (CO<sub>2</sub>)

Supercritical fluid extraction was carried out in a lab scale unit (SFE 2000, Thar Technologies Inc., Pittsburgh, PA) equipped with 1,000 mL extraction vessel. The supercritical fluid extraction system was operated by passing supercritical state  $CO_2$  (with or without co-solvent) through a fixed bed of sample particles, precipitating liquid extract in cyclone separators and finally releasing  $CO_2$  to the ambient surroundings. A high pressure pump (P-50, Thar Technologies, Inc., Pittsburgh, PA) was pre-cooled to 4 °C using a cooling heat exchanger (Fig. 1) in order to



Fig. 1 Schematic diagram of a pilot scale supercritical fluid extraction system (Developed by RCFFN and Thar Technologies Inc.)

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deliver liquid  $CO_2$  from a storage cylinder (Medigas Manitoba Ltd., Winnipeg, MB) to the extraction vessel efficiently. The purity of the  $CO_2$  used is around 99%. A second P-50 pump was used as a modifier pump to deliver co-solvent (optional) to a mixer where the co-solvent mixed with  $CO_2$  before entering into the extraction vessel.

Pressure in the extraction vessel was controlled by an automatic back pressure regulator and the pressures in the cyclone separators were maintained by a manual back pressure regulator. The back pressure regulators were usually heated to 40 °C to avoid freezing upon depressurization. The temperature of the extraction vessel and the cyclone separators was maintained by separate heaters. Thermocouples were used for instant feedback in all the temperature zones. The extraction parameter (temperature, pressure etc.) was controlled by the Thar software on the computer. The exiting fluid of CO<sub>2</sub> from the separator was expanded to the atmosphere through an outlet at the back of the separator pressure controller when the controller was opened manually at the end of the extraction process.

For each run, a total of 150 g canola press cake sample was loaded into the extraction vessel. Extraction temperature was set at 40, 50 or 60 °C. The extraction pressure was set at 300, 400 or 500 bar. The total flow rate of  $CO_2$  was set at 70 g/min.

Co-solvent was added by either pumping it from the solvent supply bottle at a certain flow rate (10%) or by pouring a certain volume (2.3%) directly into the sample extraction vessel before each extraction process (Table 1). Food-grade ethyl alcohol (Commercial Alcohols, Brenntag

Canada) was used as the co-solvent in these experiments. The composition of the ethyl alcohol used was 95% EtOH and 5% distilled  $H_2O$ . Table 1 shows the detailed extraction parameters. For each parameter, extraction was conducted in duplicate and therefore there were two extracted samples used for each analysis. For each collected sample, oil content analysis and phenolic compounds analysis were performed in tandem.

### n-Hexane Extraction of Canola Press Cake

*n*-Hexane extraction was conducted to obtain a comparison with SF-extraction. A laboratory scale Soxtec unit (Soxtec<sup>TM</sup> 2050, Foss, Eden Prairie, MN) was used to extract oil from canola press cake using *n*-hexane. Fifteen grams of ground canola press cake (sieved through 14 mesh or 1,410 µm screen) was extracted with 80 mL of *n*-hexane for 95 min. Extraction was repeated twice for each sample and then the extracted oil and defatted residues were cooled down to room temperature in the fume hood. *n*-Hexane in the oil was removed using a rotary evaporator R-205 (BÜCHI Labortechnik AG, Switzerland) and dried under nitrogen (N<sub>2</sub>) to a negligible amount and the oil was used for further studies.

#### Oil Content Analysis for SFE

The SF-extracted press cake was spread overnight in the fume hood to remove the residual ethanol prior to determination of the oil content. The initial and final oil contents of the canola press cake samples before and after

| Parameter <sup>a,b</sup> | Total flow rate<br>(CO <sub>2</sub> ) (g/min) | Co-solvent %     | Extraction vessel<br>Temperature<br>(°C) | Operating<br>pressure<br>(bar) | Cyclone separator<br>pressure (bar) | Concentration of<br>residual oil in the<br>extracted sample<br>$(g/100 \text{ g} \pm \text{SD})^{e,d}$ |
|--------------------------|---|------------------|--|--------------------------------|-------------------------------------|--|
| EP 1                     | 70  | 10 <sup>b</sup>  | 40                                       | 300                            | 60                                  | $10.3 \pm 0.8a$  |
| EP 2                     | 70  | 10 <sup>b</sup>  | 40                                       | 300                            | 70                                  | $6.7 \pm 0.1 \mathrm{b}$   |
| EP 3                     | 70  | 10 <sup>b</sup>  | 50                                       | 400                            | 50                                  | $8.2\pm0.07c$  |
| EP 4                     | 70  | 2.3 <sup>c</sup> | 50                                       | 300                            | 60                                  | $8.7\pm0.03c$  |
| EP 5                     | 70  | 2.3 <sup>c</sup> | 50                                       | 400                            | 60                                  | $2.9\pm0.2d$   |
| EP 6                     | 70  | 2.3 <sup>c</sup> | 60                                       | 400                            | 60                                  | $2.4 \pm 0.03 d$   |
| EP 7                     | 70  | 2.3 <sup>c</sup> | 50                                       | 500                            | 60                                  | $2.1\pm0.05d$  |

Table 1 Supercritical fluid extraction parameters and concentration of residual oil in the extracted cake for each sample trial

<sup>a</sup> EP, extraction parameter; the total flow rate of  $CO_2$  was kept at 70 g/min, the collector temperature was maintained at 40 °C, and the extraction time was 4 h long for all the above experimental runs

<sup>b</sup> Trials with 10% co-solvent were carried out as follows: co-solvent was run only for the first 1 h then shut off and  $CO_2$  was run for another 3 h; co-solvent 10% means a co-solvent flow rate of 8 g/min, it was calculated based on the total flow rate

 $^{c}$  Co-solvent 2.3% means 500 mL ethanol, it was calculated based on the total weight of CO<sub>2</sub> consumed in the 4 h extraction process

<sup>d</sup> The initial oil content of canola press cake was  $21.06\% \pm 0.1$  (g/100 g); the concentration of oils in the extracted sample (g/100 g)

<sup>e</sup> Values are means of four replications for each extraction parameter. Means (n = 4) with different letters are significantly different by LSD (p = 0.05)

supercritical  $CO_2$  extraction were determined using the traditional Soxhlet method (ISO/DIS 659: 2007) to compare the efficiency among the different extraction parameters. The initial oil content of the canola press cake was analyzed twice and the reported data was the mean of the two replications. To determine oil content after SFE, replications were done for each sample, and thus the reported yield was the mean of four replications for each extraction parameter. Residual oil percentages in the extracted sample with different extraction parameters were compared and statistically analyzed by least significance difference method (LSD).

# Phenolic Content of SFE Extracted Residue

The extracts were left overnight in the fume hood, evaporated the next day using the rotary evaporator, followed by drying under nitrogen (N<sub>2</sub>) to remove the residual ethanol prior to determination of phenolics. As per the oil content analysis after SFE, the reported yield for phenolic content (Table 3) was the mean of four replications of the two samples for each extraction parameter. The total phenolic content was determined by Folin-Ciocalteu's reagent method according to Swain and Hillis [20] and Schanderl [21] with some modifications. Aliquots (0.2 mL) of extracts were diluted to 0.5 mL with distilled water, and Folin-Ciocalteau's phenol reagent (0.5 mL) was added. After 3 min, 19% sodium carbonate (1 mL) was added. After 60 min, the absorption was measured at 750 nm using the DU 800 UV/Visible Spectrophotometer (Beckman Coulter Inc., Mississauga, ON, Canada). Sinapic acid (Sigma) was used for the calibration, and the results of duplicate analyses were expressed as sinapic acid equivalents (SAE). The main phenolic components were estimated according to Khattab et al. [22] using the reversed-phase DAD-HPLC (Ultimate 3000; Dionex, Sunnyvale, CA, USA). A gradient elution was performed using water/methanol (90:10) with 1.2% o-phosphoric acid as solvent A, and methanol (100%) with 0.1% o-phosphoric acid as solvent B, using C18 column (Synergi  $4\mu$ Fusion-RP 80 A°;  $150 \times 4.0$  mm 4 micron; Phenomenex, Canada). The column was maintained at 25 °C and different the flow rate was 0.8 mL/min. Chromatograms were acquired at 230, 275 and 330 nm and data were analyzed using the Chromeleon software (Version 6.8). Sample chromatograms were compared with authentic standards of sinapic acid, sinapine and sinapoyl glucose. Identity and purity of the phenolic compounds peaks were monitored through the 3d chromatograms, in addition to monitoring retention times and the symmetry of the peak. The method was validated according to the ICH guidelines [23]. Total phenolics and main phenolic compounds (sinapic acid, sinapine, sinapoyl glucose) in canola press cake residues extracted with Soxtec and SFE were compared and statistically analyzed by LSD (Table 3).

# **Results and Discussion**

Effects of SFE Parameters on Oil Yield of Canola Press Cake

In order to improve the extraction efficiency, the press cake was subjected to freeze drying and fine milling before supercritical CO<sub>2</sub> extraction. Moisture interferes with the efficiency of SFE of oil, in part by inhibiting contact between the supercritical  $CO_2$  and the sample [24]. High moisture content of the sample acts as a barrier to diffusion of supercritical CO<sub>2</sub> into the press cake as well as diffusion of oil out of the press cake [11]. Removal of water by freeze drying prior to supercritical extraction has been recommended [25]. Freeze drying and fine milling will also increase the surface area and therefore improve the diffusion of the extraction fluid through the matrix [25]. For commercial purposes, additional research will be required to determine whether supercritical fluid extraction immediately following cold pressing is as efficient as drying and fine milling of press cake prior to extraction.

Extraction yield of oil using SFE is affected by two factors: (a) the properties and stability of the substrate, and (b) the solvent effect of the supercritical fluid, which is a function of operating parameters of the SFE unit. The SFE operating parameters impact fluid density and solubility and these include pressure, temperature and choice of co-solvent [26–30]. The solvent power (and hence control of selectivity) of a supercritical fluid can be adjusted by these parameters [2].

The last column of Table 1 shows the residual oil remaining in the canola press cake following supercritical fluid extraction using the corresponding parameter combinations outlined in the table. Many combinations of temperatures, pressures,  $CO_2$  flow rates, and co-solvent flow rates were initially evaluated for extraction yield (data not shown), but the parameter combinations shown in Table 1 are indicative of the range of extraction efficiency for canola oil from press cake.

In this study, the total  $CO_2$  flow rate was held constant at 70 g/min. This was the minimum flow rate of  $CO_2$  that would adequately pressurize the extraction vessel. The press cake was soaked in the EtOH at room temperature/pressure for 1 h before the extraction process. The collection vessel temperature was also held constant at 40 °C and the run time was terminated after 4 h (Table 1). Higher oil extraction yield does not appear to be correlated with higher  $CO_2$  flow rate [31, 32]. The main impact of flow rate does appear to be a reduction in extraction time (see Table 2).

| -   | -  |   | -                                 | a                      |   |  |  |                                       |        |                   |
|---|--|---|-----------------------------------|------------------------|---|--|--|---------------------------------------|--------|-------------------|
| Method  | Materials                                      | Extraction<br>pressure<br>(bar)             | Extraction<br>temperature<br>(°C) | Extraction<br>time (h) | CO <sub>2</sub> flow<br>rate            | Mass ratio<br>(g of CO <sub>2</sub> /g<br>of sample) | Use of<br>co-solvent<br>(wt%)                | Initial oil<br>content % <sup>b</sup> | Yield% | Residual<br>oil % |
| Current RCFFN trial <sup>e</sup>  | Ground canola<br>press cake                    | 400   | 50                                | 4                      | 70 g/min                                | 112.0  | 2.3% ethanol <sup>a</sup>                    | 21.1a                                 | 18.2b  | 2.9c              |
|   | Ground canola<br>press cake                    | 500   | 50                                | 4                      | 70 g/min                                | 112.0  | 2.3% ethanol <sup>a</sup>                    |                                       | 19.0b  | 2.1c              |
| Stahl et al. [3]  | Ground   | 320   | 17                                | 3                      | N/A                                     | N/A  | None   | 40.1                                  | 38.2   | 4.5               |
|   | rapeseeds                                      | 350   | 40                                | б                      | N/A                                     | N/A  | None   |                                       | 39.3   | 3.9               |
| Dunford and   | Cooked flaked                                  | 414   | 75                                | б                      | 2.5 g/min                               | 12.9   | None   | N/A                                   | 11.4   | 32.6              |
| Temelli [11]  | canola seed                                    | 414   | 55                                | б                      | 2.5 g/min                               | 12.9   | None   |                                       | 10.9   | 33.1              |
|   | Preheated flaked                               | 414   | 75                                | ε                      | 2.5 g/min                               | 12.9   | None   |                                       | 11.1   | 29.3              |
|   | canola seed                                    | 414   | 55                                | c,                     | 2.5 g/min                               | 12.9   | None   |                                       | 8.0    | 32.4              |
| Taylor et al. [37]  | Canola flakes                                  | 680   | 80                                | 1.5                    | 5 L/min<br>(9.9 g/min) <sup>c</sup>     | 254.6  | None   | 40.5                                  | 39.8   | N/A               |
| Temelli [8]   | Canola flakes                                  | 620   | 70                                | S                      | 0.65 L/min<br>(1.29 g/min) <sup>c</sup> | 7.7  | 5 g ethanol<br>(1.3% ethanol) <sup>a.c</sup> | N/A                                   | 28     | N/A               |
|   | Canola press<br>cake                           | 620   | 70                                | S                      | 0.65 L/min<br>(1.29 g/min) <sup>c</sup> | 7.7  | 5 g ethanol<br>(1.3% ethanol) <sup>a.c</sup> |                                       | 22     | N/A               |
| Barthet<br>and Daun [38]  | Ground canola<br>seed ( <i>Brassica</i>        | 517   | 100                               | 0.5                    | 2 mL/min<br>(0.004 g/min) <sup>c</sup>  | 0.07 <sup>d</sup>                                    | None   | 47.26                                 | 40.32  | N/A               |
|   | napus)   | 517   | 100                               | 1                      | 2 mL/min<br>(0.004 g/min) <sup>c</sup>  | $0.14^{d}$   | 15% ethanol                                  |                                       | 46.27  | N/A               |
|   | Ground Canola<br>seed (Brassica                | 517   | 100                               | 0.5                    | 2 mL/min<br>(0.004 g/min) <sup>c</sup>  | 0.07 <sup>d</sup>                                    | None   | 44.03                                 | 38.32  | N/A               |
|   | rapa)  | 517   | 100                               | 1                      | 2 mL/min<br>(0.004 g/min) <sup>c</sup>  | $0.14^{d}$   | 15% ethanol                                  |                                       | 42.34  | N/A               |
| <sup>a</sup> The percentage of eth<br><sup>b</sup> The initial oil conten | anol used for our exit<br>t was measured using | periment and the<br><i>n</i> -hexane extra- | e percentage cal<br>ction         | culation of eth        | anol for Temelli [8]                    | were calculated                                      | based on the total weig                      | ght of CO <sub>2</sub>                |        |                   |

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 $^{\rm d}$  Sample weight of 2 g was used to calculate mass ratio for Barthet and Daun [38]

<sup>e</sup> Means (n = 2) without letters in common differ significantly by LSD (p = 0.05) for the initial oil content %, means (n = 4) without letters in common differ significantly by LSD (p = 0.05) for the Yield% and Residual oil%



Fig. 2 Effect of extraction pressure and temperature, collection vessel pressure and co-solvent on the yield of oil during supercritical  $CO_2$  extraction of canola press cake. The *symbols* under each *bar* indicate the actual extraction parameters. The first two digits indicate the extraction temperatures (40, 50, 60 °C); the third and fourth digits

The concentration of residual oil in the press cake was reduced with increasing extraction vessel temperature (Table 1). Setting the extraction vessel temperature to 50 °C and operating pressure to 500 bar resulted in the lowest level of residual oil (2.1 g/100 g press cake) remaining in the press cake following extraction (Table 1 Treatment EP7). Increasing the extraction vessel temperature by 10 °C (60 °C) and reducing operating pressure to 400 bar (Treatment EP6) left a slightly higher level of residual oil remaining in the press cake (Table 1).

The impact of supercritical fluid extraction parameters on yield of oil are shown in Fig. 2. Increasing extraction the vessel operating temperature resulted in higher oil yields. For example, 50 °C increased the extraction yield relative to 40 °C. As in the case of pressure, extraction yield increase with an increase in the extraction temperature for the temperature range studied. This temperature effect has been reported in several instances [17, 33–36].

The oil yield was further enhanced by increasing the pressure in the first collection vessel. To be consistent throughout, pressure in the second collection vessel was not measured, as the heater and the pressure of the second collection vessel were not turned on. No oil was collected in this vessel. The yield reported was based on the amount of extract collected in the first collection vessel only. Increasing extraction pressure increases the density of the supercritical  $CO_2$  which in turn increases its solvating power. Pressure impacts the rate of extraction and extraction time. Typically, increasing pressure increases extraction yield and reduces extraction time necessary to reach optimum yield [11, 13, 29]. It is possible that higher

indicate pressure (50, 60, 70 bar) in the first collection vessel; and the last two digits indicate the use of co-solvent (2.3 and 10%, respectively). Example: 40-60-10 = 40 °C, 60 bar, 10% co-solvent. *Bars* with *different letters* are significantly different by LSD (p = 0.05)

extraction levels could be achieved with elevated operating pressure in our study, but due to equipment limitations higher pressures could not be explored at this time.

Adding ethanol as a co-solvent (2.3%) directly to the press cake in the extraction vessel was typically more efficient than pumping 10% ethanol for 1 h into the extraction vessel (Fig. 2), especially when the operating pressure was increased from 300 to 400 bar. At 300 bar, co-solvent at 10% was more efficient. Increasing the operating pressure from 400 bar to 500 bar had very little impact on oil yield. Oil yield increased by only 3 mg/g of dried press cake. 500 bar pressure is at the upper limit of operating pressure for the Thar unit. In conclusion, optimum oil yields were obtained with extraction vessel temperatures of 50 or 60 °C, a collection vessel pressure of 60 bar and an operating pressure of 400 bar in our experiments.

Comparison of Different Supercritical CO<sub>2</sub> Oil Extraction Parameters for Canola

Table 2 provides comparative data for different operating parameters for supercritical  $CO_2$  extraction of oil from canola from published sources as compared to our results. Extraction pressure and temperature have been shown to influence oil extraction yield. Table 2 indicated that there appeared to be a trend to higher oil yield with increased extraction pressure and temperature [11, 13]. The experimental result from Barthet and Daun [38] indicated that extraction oil yield could also be improved by elevating total mass ratio.

 Table 3 Individual and total phenolic compounds of canola extracts

|                  | Phenolic compour        | ids (mg/g)              |                            |                          |                          |
|------------------|-------------------------|-------------------------|----------------------------|--------------------------|--------------------------|
|                  | Sinapic acid            | Sinapine                | Sinapoyl glucose           | Total phenolics (SAE)    |                          |
|                  |                         |                         |                            | Folic Cio                | HPLC                     |
| Soxtec extracted | $0.32 \pm 0.00$ a       | $10.39\pm0.01a$         | $1.48\pm0.02a$             | $20.96\pm0.33a$          | $13.14 \pm 0.09a$        |
| SFE 1 (EP2+)     | $0.18\pm0.00\mathrm{b}$ | $8.05\pm0.00b$          | $1.18\pm0.02b$             | $12.67\pm0.20\mathrm{b}$ | $10.10\pm0.00\mathrm{b}$ |
| SFE 2 (EP3+)     | $0.27\pm0.02\mathrm{c}$ | $7.03\pm0.00\mathrm{c}$ | $2.21 \pm 0.15 \mathrm{c}$ | $12.62\pm0.11\mathrm{b}$ | $9.48\pm0.63c$           |
| SFE 3 (EP4+)     | $0.31\pm0.00a$          | $7.88\pm0.04d$          | $2.43\pm0.01d$             | $14.15\pm0.09c$          | $10.67 \pm 0.12d$        |

Values are the means of four replications  $\pm$  standard deviation (SD), values followed by different letters in the same column are significantly different by LSD (p = 0.05)

SAE sinapic acid equivalents; Folic Cio Folin-Ciocalteu's reagent method; HPLC high performance liquid chromatography; EP extraction parameter, see Table 1 for SFE parameters of EP2, EP3, and EP4

Although the initial oil contents of samples were different in the trials listed in Table 2, the efficiency of supercritical CO<sub>2</sub> oil extraction could be evaluated by the following two aspects: (1) oil yield of SFE versus that of n-hexane extraction; (2) the residual oil left in the extracts. Most of the experimental results in Table 3 indicated that canola samples were successfully extracted with supercritical CO<sub>2</sub> at a comparable oil level to that obtained with standard *n*-hexane extraction procedure. Most of the trials shown in Table 3 did not aim at lowering residue oil level, except Stahl et al. [13] who reported low residual oil content at 3.9%. Our trials revealed that the lowest residual oil values of 2.1 and 2.9% were achieved with operating pressures of 400 and 500 bar, respectively. In addition, our extraction vessel temperatures were comparable to many previous studies and our extraction pressures were within the upper and lower limits reported in Table 2. Table 2 also indicates that the consumption of CO<sub>2</sub> in our experiments were considerably higher than most trials focused on oil extraction efficiency, which is mainly due to the high capacity of our extraction unit.

Phenolic Compounds in Supercritical CO<sub>2</sub> and *n*-Hexane Extracted Residues

Seventy percent aqueous methanolic extracts were evaluated for the retention of the phenolics in the residues of the SFE and *n*-hexane extracted press cake. Figure 3 shows the HPLC chromatogram of canola extract. To demonstrate the method specificity, the peak purity tests were performed and the diode array analysis showed that each chromatographic peak was attributable to a single component (graph B in Fig. 3). Contents of the total phenolics in canola extracts are illustrated in Table 3. It was found that sinapine (the choline ester of sinapic acid) was the major phenolic constituent in both the SF and *n*-hexane extracted canola meals and press cakes.

In addition to sinapine, residues of the press cake after SFE- and Soxtec-extraction indicated the presence of sinapic acid, sinapoyl glucose as the main phenolic compound. Four other compounds were present which could not be identified. Soxtec-extracted residues showed the retention of the highest sinapic acid, sinapine, sinapoyl glucose and total phenolic contents (mg/g) as indicated in Table 3. The SF-extracted residues showed the lowest values for these compounds. The difference in the individual compounds as well as in the total phenolic contents between the *n*-hexane and the SFE-defatted substrate could be attributed to the use of added ethanol, which is a polar organic solvent. Ethanol aided SF-extraction was capable of extracting the polar phenolic compounds as compared to *n*-hexane extraction. These results agree with previous work reported by Sun et al. [12] who found that the total phenolic contents of the  $SC-CO_2$  + ethanol-defatted canola meal were significantly  $(p \le 0.05)$  lower than those of the *n*-hexane-defatted one indicating that  $SC-CO_2$  + ethanol extraction removed 37.9% of the phenolic compounds from the meal. They also stated that the content of soluble tannins in the  $SC-CO_2$  + ethanol extract was significantly higher than the amount retained in the meal as the ethanol was able to break the protein- and carbohydrate-tannin complexes and release some tannins into the extract. Thus, ethanol enhanced the extraction of phenolics. Furthermore, hexane-extracted press cakes as compared to SFE/EtOH extracted press cakes are potential sources of the phenolic compounds. More studies are needed to examine the retention of individual phenolics using ethanol and SFE.

To conclude our findings, in the press cake substrates tested oil content could be further reduced to <4% using SFE. Retention of phenolic compounds can be influenced

**Fig. 3 a** Typical elution profile including retention times (RT) of phenolics of canola extract; **b** 3d HPLC chromatogram [*x* axis: RT, *y* axis: Absorbance (mAU)]. **a** *l* sinapoyl glucose, 2 sinapine, 4 sinapic acid, 3, 5, 6, 7 and 8 other unidentified sinapic acid derivatives



RT (min)

by the use of ethanol as a co-solvent. HPLC-DAD results indicate that sinapine, sinapic acid and sinapoyl glucose are present in the residues.

Acknowledgments Phenolic analysis was funded by the Natural Sciences and Engineering Research Council of Canada (NSERC), Canola Council of Canada and Syngenta Inc, Canada. The authors wish to acknowledge Dr. R. Khattab and L. Lin for their technical assistance.

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